TB ONT workshop – HerpeZ February 2023



Common file types and using TBProfiler

Dr Linzy Elton



We will record these sessions and put them online so you can refer back to them later on

We will also put the slides up online so you can access the notes (links and image credits)



ONT MTB Bioinformatics Pipeline



TB ONT workshop – HerpeZ February 2023





Reference Genome: MtbH37Rv_AL123456.embl



Some common file types you might encounter:

- Fast5 (.fast5) discussed in previous session
- Fastq (.fastq) discussed in previous session
- SAM (.sam)
- BAM (.bam)
- Fasta (.fna or .fasta)
- VCF (.vcf)



SAM/BAM files

SAM

- The Sequence Alignment/Map file (.sam) is a file format to save alignment information of reads mapped against reference genome sequence
- It usually starts with a header section followed by alignment information as tab separated lines for each read

BAM

- The Binary Alignment/Map file (. bam) is the compressed binary version of a SAM file that is used to represent aligned sequences
- BAM file has an index file ".bam.bai" (this file acts like an external table of contents, and allows programs to jump directly to specific parts of the bam file without reading through all the sequences)
- Not Human readable but you can open it on e.g. Artemis (a Genome Viewer by Sanger)



FASTA files

- File extension .fna or .fasta
- This file format is used for reference genomes or reference database sequences
- A FASTA file is the consensus sequence formed from a SAM/BAM file (the calculated order of most frequent residues at each position)
- In this file format, each sequence read is defined in 2 lines: header + sequence code

Sequence header/ID

>NC_015758.1 Mycobacterium tuberculosis variant africanum GM041182, complete genome TTGACCGATGACCCCGGTTCAGGCTTCACCACAGTGTGGAACGCGGTCGTCTCCGAACTTAACGGCGACC CTAAGGTTGACGACGGACCCAGCAGTGATGCTAATCTCAGCGCTCCGCTGACCCCTCAGCAAAGGGCTTG

VCF

- Variant calling format (.vcf) (you will need to unzip it if it is zipped)
- Used to identify SNPs (Single Nucleotide Polymorphism) and Insertions and/or deletions (INDELs)
- VCF files can be viewed in genome viewers or in Notepad
- Developed for large scale data storage
- You can set filter the output so you only see e.g. SNPs or insertions, as well as changing the quality threshold

61

Chnomedome

37 ##bcftools_viewVersion=1.10.2+htslib-1.10.2

- 38 ##bcftools_viewCommand=view -c1 -a -Oz -o pipeline/callers/tbprofiler_test_run/TB-profiler/
- 39 ##bcftools_viewCommand=view -v snps pipeline/callers/tbprofiler_test_run/TB-profiler/4e84cd
- 40 ##bcftools/csqVersion=1.10.2+htslib-1.10.2
- 41 ##bcftools/csqCommand=csq -p m -f /lustre/projects/CCM_shared_NGS_data/sylvia/.snakemake/com
- 42 ##INFO=<ID=BCSQ,Number=.,Type=String,Description="Haplotype-aware consequence annotation from the second second
- 43 ##FORMAT=<ID=BCSQ,Number=.,Type=Integer,Description="Bitmask of indexes to INFO/BCSQ, with 44 ##bcftools concatCommand=concat 2c672f8c-6cde-440b-9860-bfa1927f541b.vcf e8ee7b0d-7222-4b88
- 45 #CHROM POS ID REF ALT OUAL FILTER INFO FORMAT ERR1664619 DP=40;VDB=0.970148;SGB=-0.693145;MQSB=1;MQ0F=0; 46 Chromosome 6140 . G Т 225 PASS DP=46; VDB=0.682479; SGB=-0.693147; MQSB=1; MQ0F=0; Chromosome 225 PASS 47 7362 . G С 48 А 225 PASS DP=58;VDB=0.999803;SGB=-0.693147;MQSB=1;MQ0F=0; Chromosome 7582 . С DP=58;VDB=0.999706;SGB=-0.693147;MOSB=1;MOOF=0; 49 Chromosome 7585 . G С 225 PASS DP=48;VDB=0.261952;SGB=-0.693147;MOSB=1;MO0F=0; 50 Chromosome 9304 . G 225 PASS Α DP=35;VDB=0.662388;SGB=-0.693136;MQSB=1;MQ0F=0; 51 Chromosome 761155 С Т 225 PASS .
- 52 761998 . Т С 228 PASS Chromosome 53 764995 . С 225 PASS Chromosome G 781395 Т С 225 PASS 54 Chromosome . 55 1473246 . 225 PASS Chromosome Α G 56 1673425 . С Т 225 PASS Chromosome 225 PASS 1674782 . Т С 57 Chromosome 58 Chromosome 1917972 . Α G 225 PASS 59 2288868 . 225 PASS Chromosome Α С 3067464 . 225 PASS 60 Chromosome Α G

2067040 C 7

- DP=58;VDB=0.999706;SGB=-0.693147;MQSB=1;MQ0F=0;A DP=48;VDB=0.261952;SGB=-0.693147;MQSB=1;MQ0F=0;A DP=35;VDB=0.662388;SGB=-0.693147;RPB=1;MQ0F=0;A DP=56;VDB=0.159622;SGB=-0.693147;MQSB=1;MQ0F=0;A DP=54;VDB=0.998895;SGB=-0.693146;MQSB=1;MQ0F=0;A DP=43;VDB=0.7789;SGB=-0.693146;MQSB=1;MQ0F=0;A DP=31;VDB=0.846133;SGB=-0.693147;MQSB=1;MQ0F=0;A DP=39;VDB=0.923491;SGB=-0.693144;MQSB=1;MQ0F=0;A DP=37;VDB=0.860765;SGB=-0.693141;MQSB=1;MQ0F=0;A DP=35;VDB=0.788686;SGB=-0.693147;MQSB=1;MQ0F=0;A DP=49;VDB=0.35383;SGB=-0.693147;MQSB=1;MQ0F=0;A
- 225 DACC DD-42.00D-0 016064.0CD- 0 603146.00CD-1.000E-0.0

37 ##bcftools viewVersion=1.10.2+htslib-1.10.2

- 38 ##bcftools_viewCommand=view -c1 -a -Oz -o pipeline/callers/tbprofiler_test_run/TB-profiler/
- 39 ##bcftools_viewCommand=view -v snps pipeline/callers/tbprofiler_test_run/TB-profiler/4e84cd
- 40 ##bcftools/csqVersion=1.10.2+htslib-1.10.2
- 41 ##bcftools/csqCommand=csq -p m -f /lustre/projects/CCM_shared_NGS_data/sylvia/.snakemake/com
- 42 ##INFO=<ID=BCSQ,Number=.,Type=String,Description="Haplotype-aware consequence annotation from the second second
- 43 ##FORMAT=<ID=BCSQ,Number=.,Type=Integer,Description="Bitmask of indexes to INFO/BCSQ, with 44 ##bcftopls_concatCommand=concat 2c672f8c-6cde-440b-9860-bfa1927f541b.vcf e8ee7b0d-7222-4b88

45	#CHROM 1	POS	ID REF	ALT	QUA	L	FILTER	INFO) I	ORMAT	ERR1	6646	19					
46	Chromoson	me	6140		G	Т	225 PAS	S	DP=4();VDB=0	.9701	48;SC	GB=-0	.6931	45;MQ	QSB=1	; MQOF	r=0;
47	Chromoson	me	7362		G	С	225 PAS	S	DP=46	;VDB=0	.6824	79;s	GB=-0	.6931	47;MQ	QSB=1	; MQOF	-0;
48	Chromoson	me	7582		А	С	225 PAS	S	DP=58	;VDB=0	.9998	03;S0	GB=-0	.6931	47;MQ	QSB=1	;MQ0F	=0;
49	Chromoson	me	7585		G	С	225 PAS	S	DP=58	; VDB=0	.9997	06;S0	GB=-0	.6931	47;MQ	QSB=1	; MQOF	r=0;
50	Chromoson	me	9304		G	А	225 PAS	S	DP=48	; VDB=0	.2619	52;so	GB=-0	.6931	47;MQ	QSB=1	; MQOF	-0;
51	Chromoson	me	761155		С	Т	225 PAS	S	DP=35	;VDB=0	.6623	88;S	GB=-0	.6931	36 ; MÇ	QSB=1	;MQOF	=0;
52	Chromoson	me	761998		Т	С	228 PAS	S	DP=56	5;VDB=0	.1596	22;s	GB=-0	.6931	47;RI	PB=1;	MQB=1	; MQ
53	Chromoson	me	764995		С	G	225 PAS	S	DP=54	l;VDB=0	.9988	95;so	GB=-0	.6931	47;MQ	QSB=1	;MQOF	; =0;
54	Chromoson	me	781395		Т	С	225 PAS	S	DP=43	3;VDB=0	.7789	;SGB=	=-0.6	593146	; MQSI	B=1 ; M	Q0F=0	;AC
55	Chromoson	me	1473246		Α	G	225 PAS	S	DP=47	/;VDB=0	.8461	33;s(GB=-0	.6931	47;MQ	QSB=1	; MQOF	; =0;
56	Chromoson	me	1673425		С	Т	225 PAS	S	DP=31	;VDB=0	.4421	72;SC	GB=-0	.6931	1;MQS	SB=1;	MQ0F=	=0;A
57	Chromoson	me	1674782		Т	С	225 PAS	S	DP=39	;VDB=0	.9234	91;SC	GB=-0	.6931	44;MQ	QSB=1	; MQOF	; =0;
58	Chromoson	me	1917972		А	G	225 PAS	S	DP=37	/;VDB=0	.8607	65;SC	GB=-0	.6931	41;MQ	QSB=1	; MQOF	; =0;
59	Chromoson	me	2288868		А	С	225 PAS	S	DP=35	;VDB=0	.7886	86;S0	GB=-0	.6931	36 ; MÇ	QSB=1	; MQOF	; =0;
60	Chromoson	me	3067464		А	G	225 PAS	S	DP=49	;VDB=0	.3538	3;SGI	B=-0.	69314	7; MQS	SB=1;	MQ0F=	=0 ; A
61	Chromogor	mo	3067010		C	7	225 070	C	DD-41	• TZDD-0	0160	61.00	~D_ 0	6031	16.M	CD-1	· MOOT	-0.

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- 39 ##bcftools_viewCommand=view -v snps pipeline/callers/tbprofiler_test_run/TB-profiler/4e84cd
- 40 ##bcftools/csqVersion=1.10.2+htslib-1.10.2
- 41 ##bcftools/csqCommand=csq -p m -f /lustre/projects/CCM_shared_NGS_data/sylvia/.snakemake/com
- 42 ##INFO=<ID=BCSQ,Number=.,Type=String,Description="Haplotype-aware consequence annotation from the second second
- 43 ##FORMAT=<ID=BCSQ,Number=.,Type=Integer,Description="Bitmask of indexes to INFO/BCSQ, with 44 ##bcftools concatCommand=concat 2c672f8c-6cde-440b-9860-bfa1927f541b.vcf e8ee7b0d-7222-4b88

45	#CHROM POS	ID	REF	ALT	QU	AL	FILT	ER	INFO	I	FORMA	Т	ERR1	664	619							
46	Chromosome	614	0		G	Т	225	PASS	5	DP=4);VDB	8=0.	9701	48;	SGB	=-0.	693	145	; MQS	B=1	; MQ0	F=0;
47	Chromosome	7362	2		G	С	225	PASS	5	DP=4	6;VDB	8=0.	6824	79;	SGB	=-0.	693	147	; MQS	B=1;	; MQ0	F=0;
48	Chromosome	7582	2		Α	С	225	PASS	5	DP=5	B;VDB	8=0.	9998	03;	SGB	=-0.	693	147	;MQS	B=1	; MQ0	F=0;
49	Chromosome	758	5		G	С	225	PASS	5	DP=5	B;VDB	8=0.	9997	06;	SGB	=-0.	693	147	; MQS	B=1	; MQ0	F=0;
50	Chromosome	9304	4		G	А	225	PASS	5	DP=4	B;VDB	8=0.	2619	52;	SGB	=-0.	693	147	; MQS	B=1	; MQ0	F=0;
51	Chromosome	761	155		С	Т	225	PASS	5	DP=3	5;VDB	8=0.	6623	88;	SGB	=-0.	693	136	; MQS	B=1	; MQ0	F=0;
52	Chromosome	761	998		Т	С	228	PASS	5	DP=5	6;VDB	8=0.	1596	22;	SGB	=-0.	693	147	; RPB	=1;1	MQB=	1;MQ
53	Chromosome	7649	995		С	G	225	PASS	5	DP=5	4;VDB	8=0.	9988	95;	SGB	=-0.	693	147	; MQS	B=1	; MQ0	F=0;
54	Chromosome	7813	395		Т	С	225	PASS	5	DP=4	3;VDB	8=0.	7789	; SO	6B=-	0.69	93140	б ;М	2SB=	1;M(20F=	0;AC
55	Chromosome	1473	3246		Α	G	225	PASS	5	DP=4	7;VDB	B=0.	8461	33;	SGB	=-0.	693	147	; MQS	B=1;	; MQ0	F=0;
56	Chromosome	1673	3425		С	Т	225	PASS	5	DP=3	l;VDB	8=0.	4421	72;	SGB	=-0.	693	11;1	MQSB	=1;1	MQOF	=0;A
57	Chromosome	1674	4782		Т	С	225	PASS	5	DP=3	9;VDB	B=0.	9234	91;	SGB	=-0.	693	144	; MQS	B=1	; MQ0	F=0;
58	Chromosome	191	7972		А	G	225	PASS	5	DP=3	7;VDB	B=0.	8607	65;	SGB	=-0.	693	141	; MQS	B=1;	; MQ0	F=0;
59	Chromosome	2288	8868		Α	С	225	PASS	5	DP=3	5;VDB	B=0.	7886	86;	SGB	=-0.	693	136	; MQS	B=1;	; MQ0	F=0;
60	Chromosome	306	7464		Α	G	225	PASS	5	DP=4	9;VDB	3=0.	3538	3;5	GB=-	-0.6	59314	47;1	MQSB	=1;1	MQOF	=0;A
61	Chromogomo	306	70/0		C	7	225					~ -0	0160	61.	CCD.	- 0	603	116	MOC	D-1	• MOO	$\mathbf{r} = 0$.

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45	#CHROM POS	ID REF	ALT	QUA	L	FILTER I	NFO FORMAT ERR1664619
46	Chromosome	6140	•	G	Т	225 PASS	<pre>DP=40;VDB=0.970148;SGB=-0.693145;MQSB=1;MQ0F=0;</pre>
47	Chromosome	7362		G	С	225 PASS	<pre>DP=46;VDB=0.682479;SGB=-0.693147;MQSB=1;MQ0F=0;</pre>
48	Chromosome	7582		А	С	225 PASS	<pre>DP=58;VDB=0.999803;SGB=-0.693147;MQSB=1;MQ0F=0;</pre>
49	Chromosome	7585		G	С	225 PASS	<pre>DP=58;VDB=0.999706;SGB=-0.693147;MQSB=1;MQ0F=0;</pre>
50	Chromosome	9304		G	А	225 PASS	<pre>DP=48;VDB=0.261952;SGB=-0.693147;MQSB=1;MQ0F=0;</pre>
51	Chromosome	761155		С	Т	225 PASS	<pre>DP=35;VDB=0.662388;SGB=-0.693136;MQSB=1;MQ0F=0;</pre>
52	Chromosome	761998		Т	С	228 PASS	<pre>DP=56;VDB=0.159622;SGB=-0.693147;RPB=1;MQB=1;MQ</pre>
53	Chromosome	764995		С	G	225 PASS	<pre>DP=54;VDB=0.998895;SGB=-0.693147;MQSB=1;MQ0F=0;</pre>
54	Chromosome	781395		Т	С	225 PASS	DP=43;VDB=0.7789;SGB=-0.693146;MQSB=1;MQ0F=0;AC
55	Chromosome	1473246		А	G	225 PASS	DP=47;VDB=0.846133;SGB=-0.693147;MQSB=1;MQ0F=0;
56	Chromosome	1673425		С	Т	225 PASS	DP=31;VDB=0.442172;SGB=-0.69311;MQSB=1;MQ0F=0;A
57	Chromosome	1674782		Т	С	225 PASS	<pre>DP=39;VDB=0.923491;SGB=-0.693144;MQSB=1;MQ0F=0;</pre>
58	Chromosome	1917972		А	G	225 PASS	<pre>DP=37;VDB=0.860765;SGB=-0.693141;MQSB=1;MQ0F=0;</pre>
59	Chromosome	2288868		А	С	225 PASS	<pre>DP=35;VDB=0.788686;SGB=-0.693136;MQSB=1;MQ0F=0;</pre>
60	Chromosome	3067464		А	G	225 PASS	DP=49;VDB=0.35383;SGB=-0.693147;MQSB=1;MQ0F=0;A
61	Chromogomo	3067040		C	7		$DD = 42 \cdot V DD = 0$ 916964 · SCD = 0 693146 · MOSD = 1 · MODE = 0 ·

t file	length : 15,069 lines : 72	Ln:1 Col:1 Pos:1	Unix (LF) UTF-	8 INS
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TBProfiler

- Can detect drug resistance-conferring mutations and predict drug-resistant phenotypes
- Two versions: a browser-based one and a command line one (you will need Linux or MacOS for this)
- The pipeline is rapidly developing so make sure you use the latest version (currently v.4.4.1)
- Works with Nanopore data and paired reads from Illumina
- Analysis pipelines is pretty much standardised; the database is manually curated database hosted on separate repository constantly updated, based on emergence of new evidence for the inclusion or exclusion of mutations affect the Accuracy of the AMR prediction

Important! The mutations and resulting library files are in reference to the H37Rv (NC_000962.3/AL123456.3) reference genome

TBProfiler versions

Web-	based	Command line				
Pros	Cons	Pros	Cons			
Simple to use	Limit on the size of the files is 4GB	Once downloaded, doesn't require internet	Linux or MacOS only			
User friendly, interactive interface	Requires stable internet connection	Fast (although depends on computer power)	Must understand line command to use			
Creates a browser- based, easy to interpret output file		Unlimited file size	Output .json file is less easy to interpret			

https://tbdr.lshtm.ac.uk/

TB-Profiler Home Upload SRA data

TB Profiler

Welcome to the webserver of TB-Profiler - a pipeline which allows users to analyse *M. tuberculosis* whole genome sequencing data to predict lineage and drug resistance. Follow the instructions below to upload a new sample or view analysed runs.

How does it work?

The pipeline searches for small variants and big deletions associated with drug resistance. It will also report the lineage. By default it uses Trimmomatic to trim the reads, BWA (or minimap2 for nanopore) to align to the reference genome and GATK (open source v4) to call variants.

TB-Profiler Home Upload SRA data

Upload your next generation sequencing data in **fastQ** format. Files will be processed using the tb-profiler pipeline with default parameters. You may select the technology used to generate the data (Illumina or Oxford Nanopore). Samples will be process with a first in first out policy so please be patient as there may be runs waiting to be processed before yours.

Please note: at the moment we can only accomodate uploads of files under 4GB. If you have files which are larger or you require many isolates to be processed and have access to a linux or osX operating system then it might be worthwhile to run the commandline version of tb-profiler. For more information on this please visit the github repository.

Batch upload You can upload multiple samples together to batch process them. By default, paired files must use _1.fastq.gz and _2.fastq.gz as the file suffix in order for them to be paired correctly. The suffix can be changed in the advanced options. Drop files here to upload

Drug class definitions

Samples can be classed into different types using the following definitions

Туре	Drugs resistance
Sensitive	No drug resistance
Pre-MDR	Rifampicin or isonisazid
MDR	Rifampicin and isoniazid
Pre-XDR	MDR and any fluoriquinolone
XDR	MDR and (any fluoriquinolone and any group A drug)
Other	Resistance to any drug but none of the above categories

Mutation format

Mutations must follow the HGVS nomenclature. Information on this format can be found here. The following types of mutations are currently allowed:

- Amino acid substitutions. *Example: S450L in rpoB would be p.Ser450Leu*
- Deletions in genes. Example: Deletion of nucleotide 758 in tlyA would be c.758del
- Insertion in genes. Example: Insertion of GT between nucleotide 1850 and 1851 in katG would be c.1850_1851insGT
- SNPs in non-coding RNAs. Example: A to G at position 1401 in rrs would be r.1401a>g
- SNPs in gene promoters. Example: A to G 7 bases 5' of the start codon in pncA c.-7A>G

Important! The mutations and resulting library files are in reference to the H37Rv (NC_000962.3/AL123456.3) reference genome

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Let's have a look at TB-profiler output reports

Browser-based report (768345cd-b4b1-4d22-a41b-2881dcdf3af3)

TB-Profiler Home Upload SRA data

TB-Profiler result

Run: 768345cd-b4b1-4d22-a41b-2881dcdf3af3

	Summary
Run ID: 768345cd-b4b1-4d22-a41b-2881dcdf3af3	
Sample name:	
Date: 26-01-2023 15:39:34	
Number of reads: 30203	
Percentage reads mapped: 95.41	
Strain: lineage2.2.2	
Spoligotype: 000000000000000	
Drug-resistance: Pre-XDR-TB	

тв о	Drug	Resistance	Supporting mutations
R	Rifampicin	R	rpoB p.Ser450Leu (0.91)
	Isoniazid		
	Ethambutol	R	embB p.Met306lle (1.00)
	Pyrazinamide	R	pncA p.Cys14Arg (1.00)
	Streptomycin	R	gid p.Leu79Ser (1.00)
	Fluoroquinolones	R	gyrA p.Asp94Gly (1.00)
	Moxifloxacin	R	gyrA p.Asp94Gly (1.00)
	Ofloxacin	R	gyrA p.Asp94Gly (1.00)
	Levofloxacin	R	gyrA p.Asp94Gly (1.00)

Lineage Table: The lineage is inferred by analysing lineage specific SNPs

Lineage	Family	Main Spoligotype	RDs	Frequency
lineage2	East-Asian	Beijing	RD105	1.0
lineage2.2	East-Asian (Beijing)	Beijing-RD207	RD105;RD207	1.0
lineage2.2.2	East-Asian (Beijing)	Beijing-RD105/RD207	RD105;RD207	1.0

Drug resistance-Associated Mutations: This table reports mutations found in candidate resistance genes which have been associated with drug resistance

	Chromosome			Estimated	
Gene	position	Mutation	Туре	fraction	Drugs

TB ONT workshop -

JSON report

▶ 0: ▶ 1: ▶ 2: ▼ 3: lin: family: spoligotype: rd: frac: main lin: sublin: variants: **v** 0: chrom: genome_pos: ref: alt: freq:

v lineage:

feature_id: type: nucleotide_change: protein_change:

v annotation: ▼ 0:

type:

drug:

{...} {...} {...}

"lineage4.1.2.1" "Euro-American (Haarlem)" "T1;H1" "RD182" 0.9984939759036144

"lineage4" "lineage4.1.2.1"

"Chromosome" 761155 "C" "T" 1 "CCP43410.1" "missense_variant" "c.1349C>T" "p.Ser450Leu"

"resistance_association_confidence" "rifampicin"

sublin:	"lineage2.2.2"
▼ dr_variants:	
▶ 0:	{}
▼ 1:	
chrom:	"Chromosome"
genome_pos:	761155
ref:	"C"
alt:	"Т"
depth:	11
freq:	0.909090909090909091
feature_id:	"CCP43410"
type:	"missense_variant"
<pre>nucleotide_change:</pre>	"c.1349C>T"
protein_change:	"p.Ser450Leu"
▶ annotation:	[]
alternate_consequences	[]
change:	"p.Ser450Leu"
locus_tag:	"Rv0667"
gene:	"rpoB"
drugs:	[]
<pre></pre>	
0:	"rifampicin"
▶ 2:	\ 5
▶ 3:	{}
▶ 4:	{}
<pre>> other_variants:</pre>	[]
drtype:	"Pre-XDR-TB"

TXT report

TBProfiler report

The following report has been generated by TBProfiler.

Summary

ID: pan346 Date: Fri Feb 4 14:16:27 2022 Strain: lineage4.1.2.1

Drug-resistance: MDR

Lineage report

Lineage Estimate	d Fract	ion	Family	/ Spoligo	type	Rd
lineage4	1.000	Euro-Am	erican	LAM;T;S	;X;H	None
lineage4.1	1.000	Euro-Am	erican	T;X;H	None	
lineage4.1.2	0.998	Euro-Am	erican	T;H	None	
lineage4.1.2.1	9.998	Euro-Am	erican	(Haarlem)	T1;H1	RD182

Resistance report

Drug	Genotypic	Resistance	Mutations
Rifampicin R		rpoB	p.Ser450Leu (1.00)
Isoniazid		katG	p.Ser315Thr (1.00)
Ethambutol		embB	p.Met306Ile (1.00)
Pyrazinamide		pncA	p.Leu85Pro (1.00)
Strepto	omycin R	rpsL	p.Lys43Arg (1.00)
Fluoroquinolones			
Amikacin			
Kanamyo	in		

Tree scale: 0.00001 🖂

PARA-AMINOSALISYLIC_ACID Lineage **FLUOROQUINOLONES** AMINOGLYCOSIDES Lineage1 STREPTOMYCIN PYRAZINAMIDE CAPREOMYCIN ETHIONAMIDE Lineage2 ETHAMBUTOL BEDAQUILINE CLOFAZIMINE KANAMYCIN RIFAMPICIN LINEZOLID SONIAZID AMIKACIN Lineage3 Lineage4 Lineage5 ERR551732 Lineage6 ERR553000 ERR1679637 Lineage7 ERR1679623 ERR1034610 Bovis ERR046855 Other ERR181983 ERR400440 ERR1035363 ERR040132 Drug resistance ERR036228 Sensitive SRR1162491 ERR552937 Drug-resisant ERR551751 ERR040133 MDR ERR1034587 XDR

TB ONT workshop – HerpeZ February 2023

