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Agility Program Biweekly Progress

Agility Program: To enable the rapid assessment of the biological impacts of new variants of SARS-CoV-2

Partners: UK Health Security Agency (UKHSA – formerly Public Health England) National Institute for Biological Standards and Control (NIBSC)







Slideset provided on a biweekly basis to update latest in vitro neutralization activity and in vivo pathogenesis and cross protection data against SARS-CoV-2 virus variants

Find this slide set posted at:

https://epi.tghn.org/covax-overview/enabling-sciences/agility_epi/#ref1

Variants of Concern and Interest Monitored by the Agility Project

| WHO Variants of Concern Status* | | Deselected Variants | |
|---|--|---|------------------------|
| Delta - B.1.617.2 | Assessed ² | Tested within Agility | Not tested with |
| Omicron | | Alpha – B.1.1.7 | Eta (B.1.525) |
| BA.1 | Assessed ² | Beta – B.1.351 | Epsilon (B.1.427/B.1.4 |
| BA.1.1 BA.2 | Assessed ² Assessed ² Sourced Sourced | Gamma – P.1 | Theta (P.3) |
| BA.2.12.1 | | Zeta (P.2) – sourced from Fiocruz | lota (B.1.526+E484K) |
| BA.3 | | Zeta (P.2) – sourced from BEI | |
| BA.4 BA.5 | Seeking Sourced | Карра (В.1.617.1) | |
| Superscripts denote assessed at 1 or 2 sites in vitro | | – Mu (B.1.621) | |
| | | Alpha + E484K | |
| Recombinants | Status* | Lambda (C.37) (single lab | |
| Omicron x Delta recombinant (XD) | Seeking | evaluation) | |
| Omicron x Delta recombinant (XF) | Sourced | AY.1 | |
| Alpha x Delta recombinant (XC) | Sourced | AY.4.2 | |
| BA.1 x BA.2 recombinant (XE) | Sourced | *C.1.2 ^a (single lab evaluation) | |
| | | *Isolate D190^b (single lab evaluation) | |

*Isolates provided by Alex Sigal, African Health Research Institute, pursued for reasons of interesting Spike mutations

^ahttps://www.medrxiv.org/content/10.1101/2021.08.20.21262342v1 ^bhttps://www.medrxiv.org/content/10.1101/2021.09.14.21263564v1.full

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Table updated as of May 27, 2022

Agility Project: Variant Growth/Testing for Neutralization Phenotype

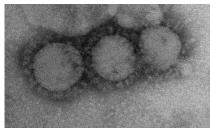
| | Va | riant | Sourcing or Propagation Seeking/In progress/Complete | Characterisation | In vitro (neutralisation) In progress/Complete/No longer required | In vivo Not selected/Planning/in progress/In-life complete |
|-------|------------------------------|-------------------------------------|---|------------------|--|---|
| WHO | [†] Alpha (B.1.1.7) | | Complete | Complete | Complete | In-life complete |
| VOCs | [†] Beta (B.1.351) | | Complete | Complete | Complete | |
| | [†] Gamma (P.1) | | Complete | Complete | Complete | |
| | Delta (B.1.617.2) | | Complete | Complete | Complete | In-life complete – reporting underway |
| | | BA.1 | Complete | Complete | Complete | In-life complete –manuscript link ^a |
| | | BA.1.1 | Complete | Complete | Complete | |
| | | BA.2 | | Complete | Complete | |
| | Omicron (B.1.1.529) | BA.2.12.1 | | | | |
| | | BA.3 | Sourced | In progress | | |
| | | BA.4 | Seeking | | | |
| | | BA.5 | Sourced | | | |
| | | Omicron x Delta recombinant (XD) | Seeking | | | |
| Other | Recombinants | Omicron x Delta recombinant (XF) | Sourced | In progress | In progress | |
| Other | Recompinants | Alpha x Delta recombinant (XC) | Sourced | In progress | In progress | |
| | | BA.1 x BA.2 recombinant (XE) | Sourced | | | |

[†]No longer a WHO VOC





Wildtype virus Quality Control



- Most viruses isolated from clinical material through UKHSA's network
- Some have been isolated elsewhere and donated by other institutes
 - G2P consortium
 - Barclay 'flu lab (Imperial College, London)
 - Oxford University, UK
 - Fiocruz, Brazil
 - Sheba Medical Centre, Israel
 - AHRI, South Africa
- All are grown into working banks and quality control assessments are performed
 - CoAs issued

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• Virus stocks available from NIBSC and EVAg

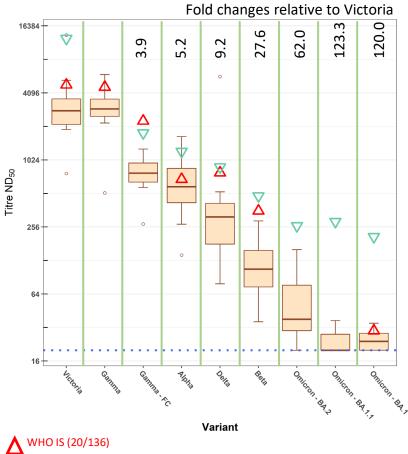
| Criteria | Result |
|--|---|
| Passage history, cell line(s) used, MOI and harvest details | Recorded |
| Morphology | Transmission electron microscopy |
| Cytopathic effect | Record appearance |
| Viable titre | Plaque forming units on Vero E6 (and additionally/alternatively VAT or foci) |
| Usage dilution in micro- neutralisation assay (MNA) | For ~130 focus forming units/well in non- neutralisation control |
| Sterility | 7 days in TSB & Thioglycollate at 22° and 37°C $$ |
| Absence of mycoplasma | ECACC validation PCR test |
| Sequence analysis – Nanopore/Arctic v3 | Confirm presence of furin cleavage site, identity, lineage (<i>fast</i>) |
| Sequence analysis – Illumina NGS/SISPA | Examination of minor variants, absence of contaminants, fill in any 'missed' regions due to Arctic protocol primer mismatches (<i>detailed</i>) |

Questions? Reach us at agility@cepi.net

Table updated as of May 27, 2022

Live-virus in vitro antibody neutralization assay progress



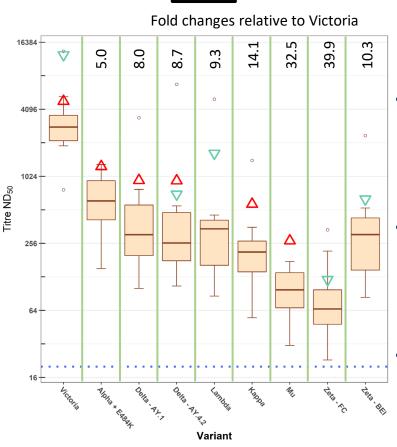


VNIBSC WS (21/234)









C.1.2 and D190 provisional data (presented in earlier updates) has been removed because subsequent WGS analysis revealed that these virus banks did not meet the quality standards specified by the Agility programme.

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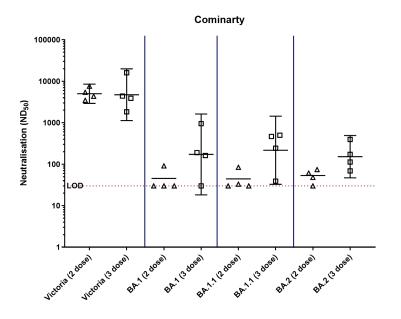
- UKHSA and NIBSC neutralisation assays behave comparably across variants
 - Data are presented as geometric mean of titres from both labs
 - Variants shows various degree of resistance to the panel - only statistically significant (p<0.001) fold changes relative to Victoria are shown)
 - IS & WS generally show the least neutralisation reduction compared to individual samples – can't be used to correct ND_{50} s across variants
 - Omicron exhibit the largest drops in ND_{50} seen to date
 - Many of the titres below assay limit of detection for BA.x variants which is likely to make this fold-change an underestimate

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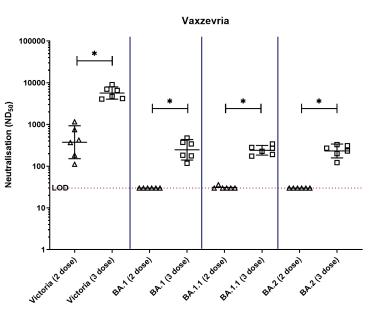
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Cross-neutralisation results from the UKHSA ESCAPE staff vaccinee study

* p<0.05 in paired t test



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Red line indicates assay limit of detection (LOD) * Many titres below LOD for BA.x variants which is likely to make fold-change an underestimate. Percentage below LOD indicated

• Serum samples from vaccinated UKHSA staff volunteers following 2 or 3 vaccinations

- Cominarty/ Vaxzevria refers to the primary course (first 2 doses). All staff received an mRNA booster vaccine.
- Group sizes: Cominarty n=4; Vaxzevria n=6
- All sera from boosted individuals display comparable titres regardless of whether initial course was Cominarty (RNA) or Vaxzevria (ChAdOx).
- All boosted individuals display improvements in titre including to the Omicron variant
- Boosted individuals who received Vaxzevria for their primary course display significant increases in titre relative to their initial course titres
 - This effect is smaller for those who received a Cominarty primary course due to the higher initial titres after two doses

https://manchestercrf.nihr.ac.uk/about/our-impact-case-studies/escape/

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The broader scientific community is currently collecting biological infection data to understand disease severity and immune reponse to variants of concern in the following ways, plus many others:

- Human clinical studies assessing vaccine effectiveness against variant infections
- Animal studies in various laboratory model species to evaluate effectiveness of original vaccines against variants, and new vaccines, need for boosters, etc.

The Agility Program is leveraging CEPI Preclinical Laboratory Network Partners to perform hamster modeling studies under high ethical standards

- CEPI Network of Partners was established in 2019 via a call for proposals to engage laboratories with high animal ethics standards, biocontainment laboratory capabilities and high-quality research methods that meet regulatory requirements
- All animal studies are performed in accordance with UK NC3Rs guidelines (<u>https://www.nc3rs.org.uk/the-3rs</u>)
- All research is done in compliance with CEPI's <u>Animals in Research Policy</u>







Primary infection studies confirmed typical coronavirus disease; and Re-Infection Studies showed solid protection from disease in hamsters, even across variants

| Initial Infection | Re- infection | Clinical signs after re-infection? | Weight loss after re-infection? | Protection against re- infection? |
|----------------------|------------------|---------------------------------------|------------------------------------|--------------------------------------|
| Alpha | Delta | No | No | Yes |
| Victoria | Delta | No | No | Yes |
| Beta | Gamma | No | No | Yes |
| Beta | Beta | No | No | Yes |
| Gamma | Beta | No | No | Yes |
| Gamma | Gamma | No | No | Yes |
| Victoria | Mu | No | No | Yes |
| Victoria | Zeta | No | No | Yes |
| Victoria | Omicron | No | No | Yes* |

All studies were conducted in compliance to all UK government regulatory requirements. In-life phase complete: full data analysis is underway, with ELISA, microneutralization and pathology data pending. *pre-print released Dec 24 https://www.biorxiv.org/content/10.1101/2021.12.24.474081v1



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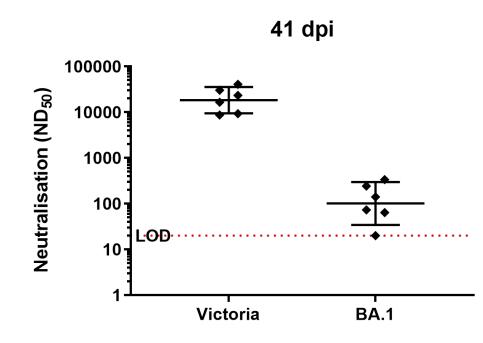
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- ✓ For all VOCs tested, prior infection was able to protect against secondary infection 28 days later.
- None of the combinations of VOCs tested showed escape from immunity.
- Preliminary pathology data has not identified any difference between VOCs with the exception of **Omicron for which similar** lesions in the lung and upper respiratory tract were present, but with lower severity.

Table updated as of May 27, 2022

Variant assessment – In vivo

Preliminary data



Hamsters infected with Vic and sera taken 41 days pi

Similar fold-change to human convalescent sera with a 150fold drop

Omicron neutralisation titres above the LOD

Ryan et al 2021, Convalescence from prototype SARS-CoV-2 protects Syrian hamsters from disease caused by the Omicron variant https://www.biorxiv.org/content/10.1101/2021.12.24.474081v1

Important considerations for laboratory methods

- Serial propagation of SARS-CoV-2 variants in Vero E6 or other cell types may lead to furin cleavage site mutations that affect how the virus grows and behaves in vitro or in vivo. Propagation of unwanted mutations can be mitigated by growth in cells such as Vero/hSLAM and by frequent sequence confirmation (deep sequence methods preferred). <u>link</u>
- <u>NIBSC Working Standard</u> should be used for neutralization assays, but it performs differently for each variant. Any data presented comparing the WHO IS should always identify the variant under test.

Recent relevant publications

- <u>Quantification of SARS-CoV-2 neutralizing antibody by wild-type plaque reduction neutralization</u>, <u>microneutralization and pseudotyped virus neutralization assays</u> Nature Protocols **16**, 3114-3140 (2021)
- <u>A cautionary perspective regarding the isolation and serial propagation of SARS-CoV-2 in Vero cells</u> NPJ Vaccines **6**:83 (2021)

Recent online conference presentations

- 13 January 2022: WHO Animal Models Working Group meeting
- 22 Feb 2022: Joint ECDC and WHO lab assay working group meeting
- 17 March 2022: New Variant Assessment Platform (NVAP) module on SARS-CoV-2 Risk Assessment and Virology <u>https://www.gov.uk/guidance/new-variant-assessment-platform</u>



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