

The background of the slide is a dark blue field filled with various microscopic images of viruses. Some are spherical with distinct spikes (like coronaviruses), while others are more complex, rod-like, or have different surface textures. The lighting is dramatic, with some viruses appearing to glow or be highlighted against the dark background.

The INHALE Project: What have we learnt about the molecular diagnostics of HAP/VAP?

Dr Vicky Enne

University College London

Twitter: @Beat_AMR_Bugs

@HAP_Diagnostics

Website: www.ucl.ac.uk/inhale-project

The logo for the INHALE project. The word 'INHALE' is written in a large, bold, black, sans-serif font. The letter 'H' is replaced by a stylized graphic of a DNA double helix. The two strands of the helix are colored: the left strand is blue and the right strand is red, with horizontal rungs connecting them in yellow, green, and blue.



Sample arrives
in lab



Microscopy/Gram
Stain
(a few min)



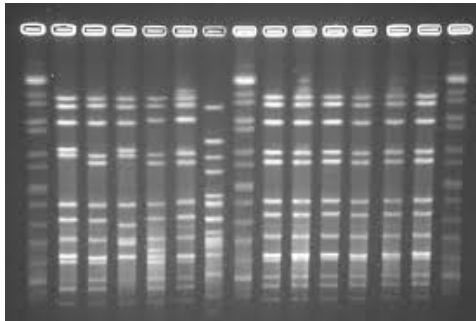
Culture
(16h-48h)



Identify growth: MALDI-
TOF/Biochemical testing
(a few min – 24h)

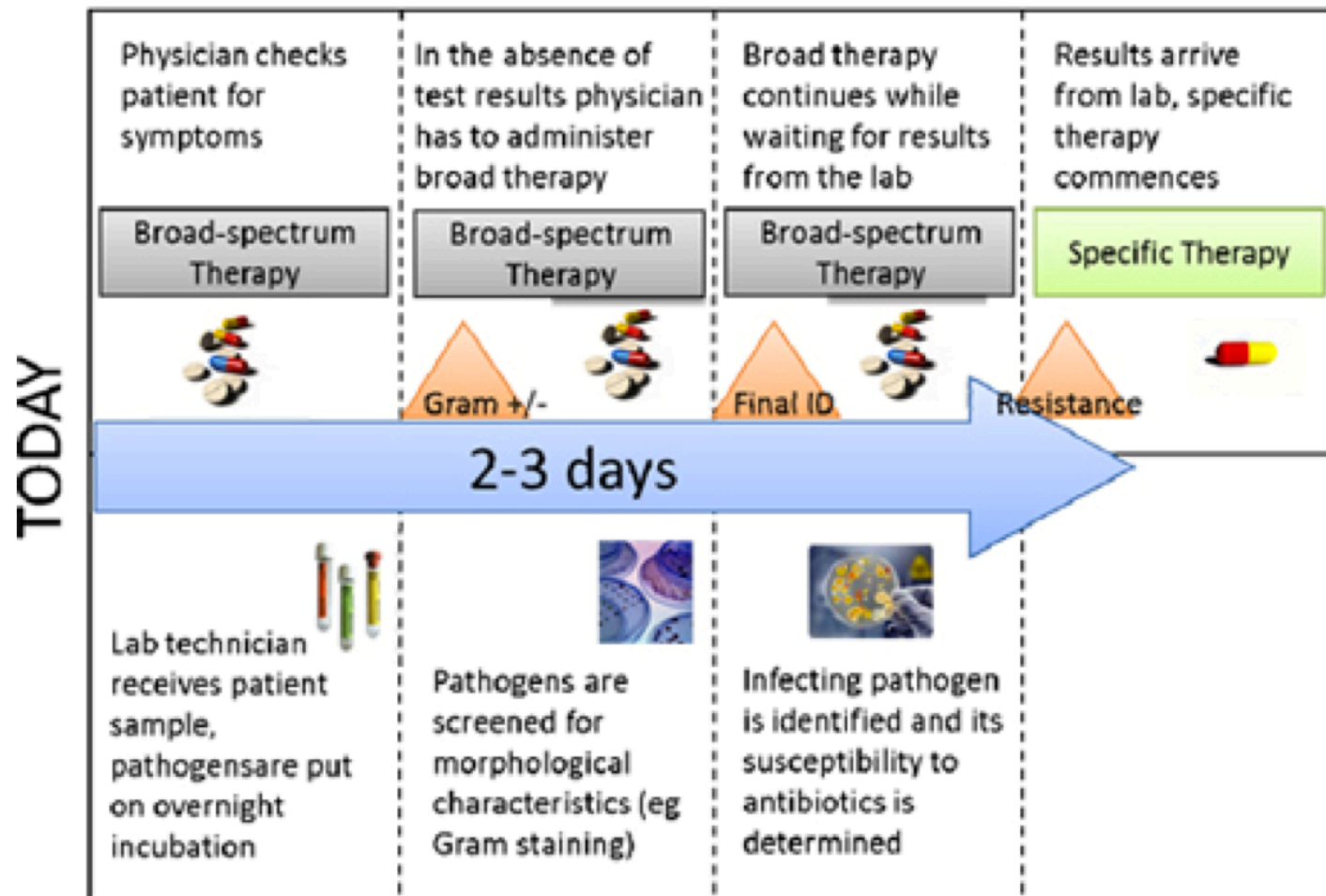


Antimicrobial susceptibility
testing
(24-48h)



Further testing /
reference laboratory
(up to 2 weeks)

The Need for Rapid Diagnostics



Source : Bionest analysis

What Factors Contribute to the Adoption of Molecular Diagnostics?

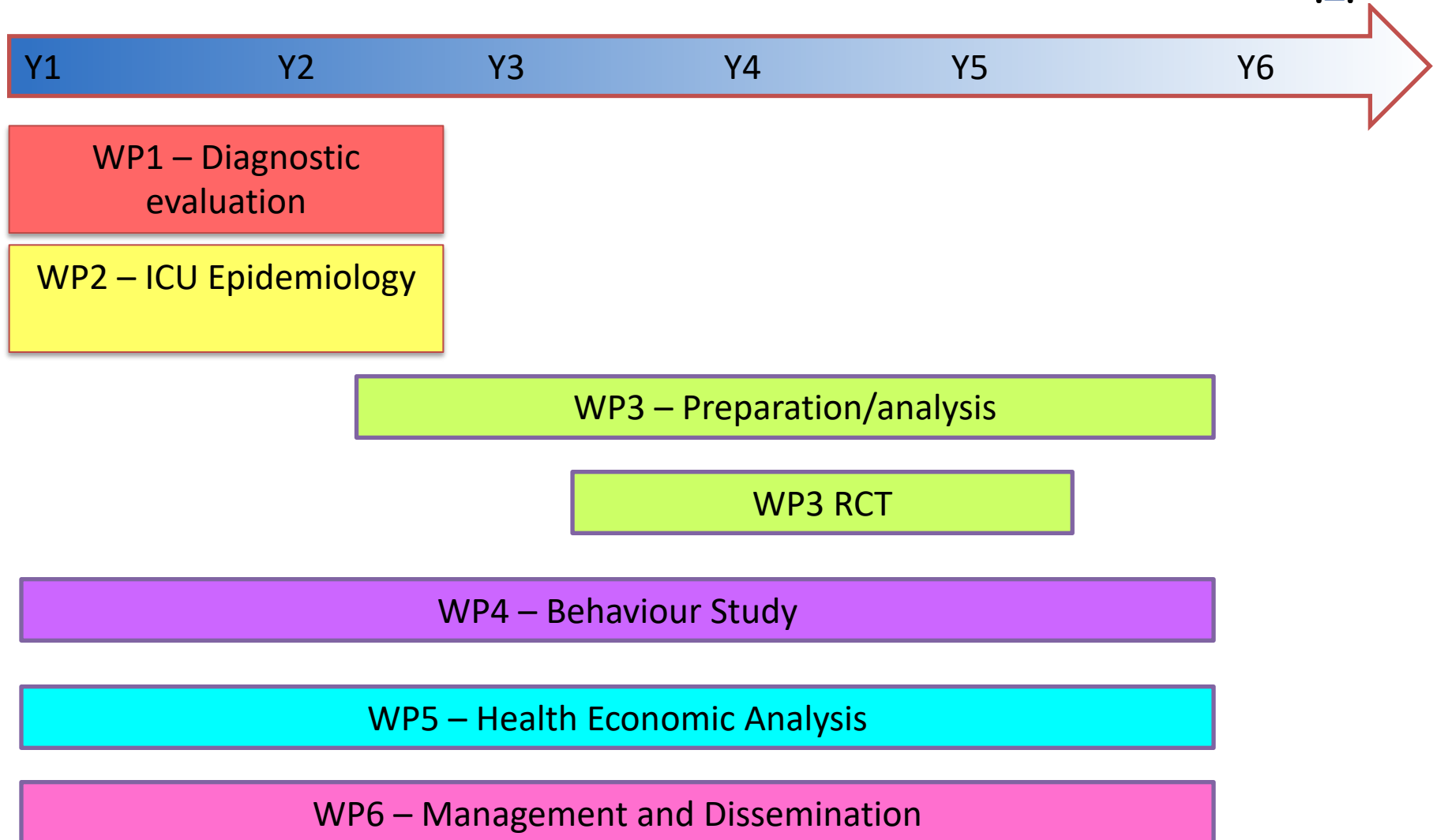
In the Clinical Laboratory

- Can it be integrated into the clinical pathway/laboratory workflow?
 - How much does it cost – **who pays?**
 - Is it easy to use, how much training is required?
 - How much space is needed?
 - How long does it take?
- Is the machine reliable
 - How often does it break down?
 - Is the customer support adequate?

At the Bedside

- Are the results accurate
 - Does the test do “what it says on the tin”?
 - Does it look for the important pathogens/resistance genes?
- Do doctors trust the results & know what to do with them?
- Does using the test actually make a difference:
 - To levels of antibiotic use?
 - To patient outcomes?

INHALE Programme Outline



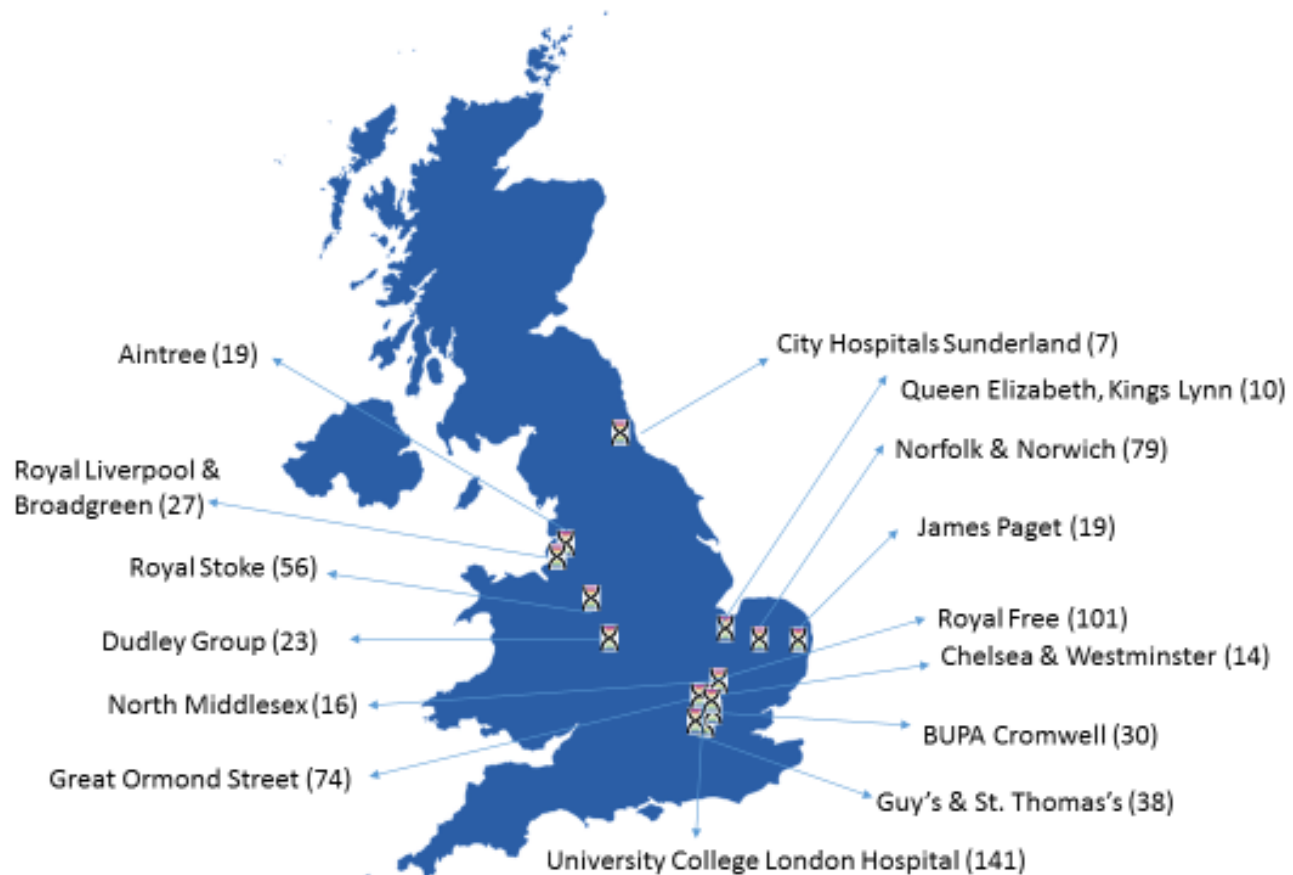
Hospital-acquired Pneumonia (HAP)

- Defined as pneumonia that occurs >48h after hospital admission
 - VAP occurs in ventilated patients
- 1.5% of inpatients in UK
- Approx. 200k patients/year
- Mortality rate for HAP/VAP is approx. 25-50%
- Increases to 75% if MDR pathogen
- HAP/VAP adds approx. 8 days to ICU stay
 - costs an additional \$30,000 - \$37,000 per patient



Project Participants

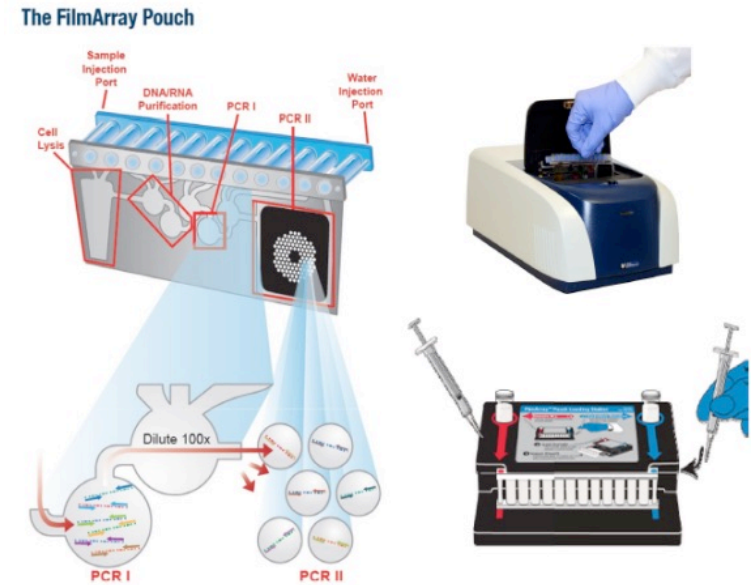
- UCLH/UCL/UEA (Lead organisations)
- 15 Critical Care Units in England representing diverse case-mix



INHALE WP1: Head to Head Comparison of 3 Rapid Molecular Tests for Pneumonia Diagnosis vs. Routine Microbiology Culture



Curetis Unyvero Pneumonia Test



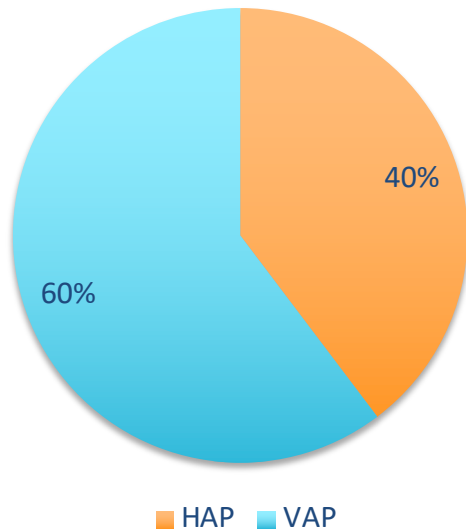
Biofire FilmArray Pneumonia Panel



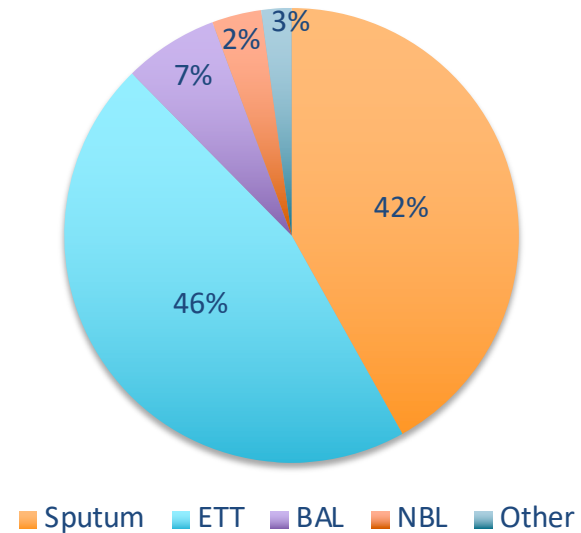
Oxford Nanopore Technologies MinION metagenomic sequencing

Clinical Evaluation Sample Characteristics

Type of Pneumonia



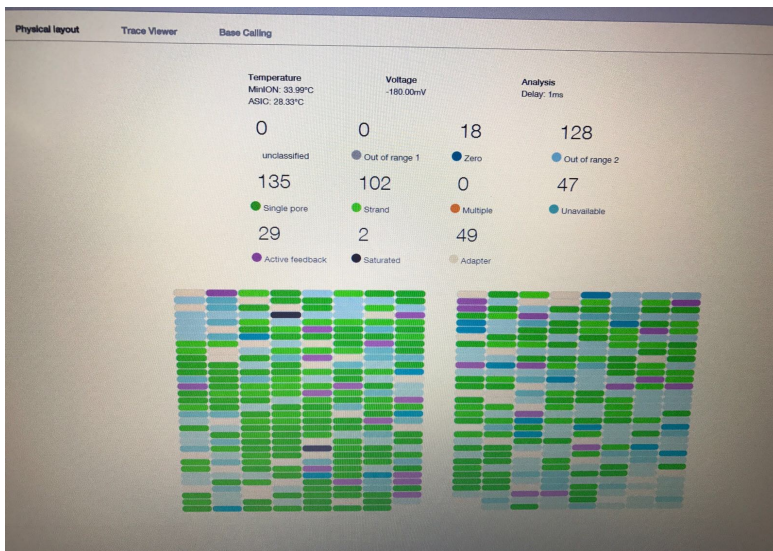
Sample Type



- 654 eligible samples collected from 15 hospitals

Oxford Nanopore MinION NGS Diagnostics

- Rapid, low-cost NGS sequencing based on nanopore technology
- Capability to obtain full organism & resistance gene profile – not limited to selected targets

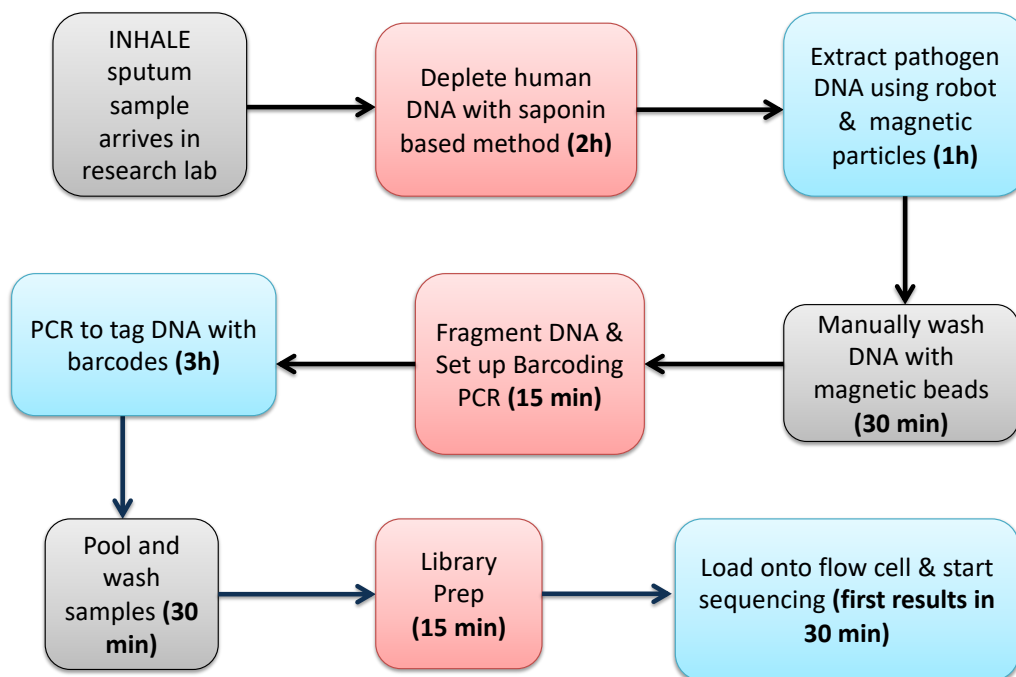


- World first comprehensive trial of *rapid* metagenomics for the diagnosis of infection (336 specimens over 9 months)



INHALE Laboratory Custom Work Flow for MinION

Processing: Total time to Result = approx. 7h



nature
biotechnology

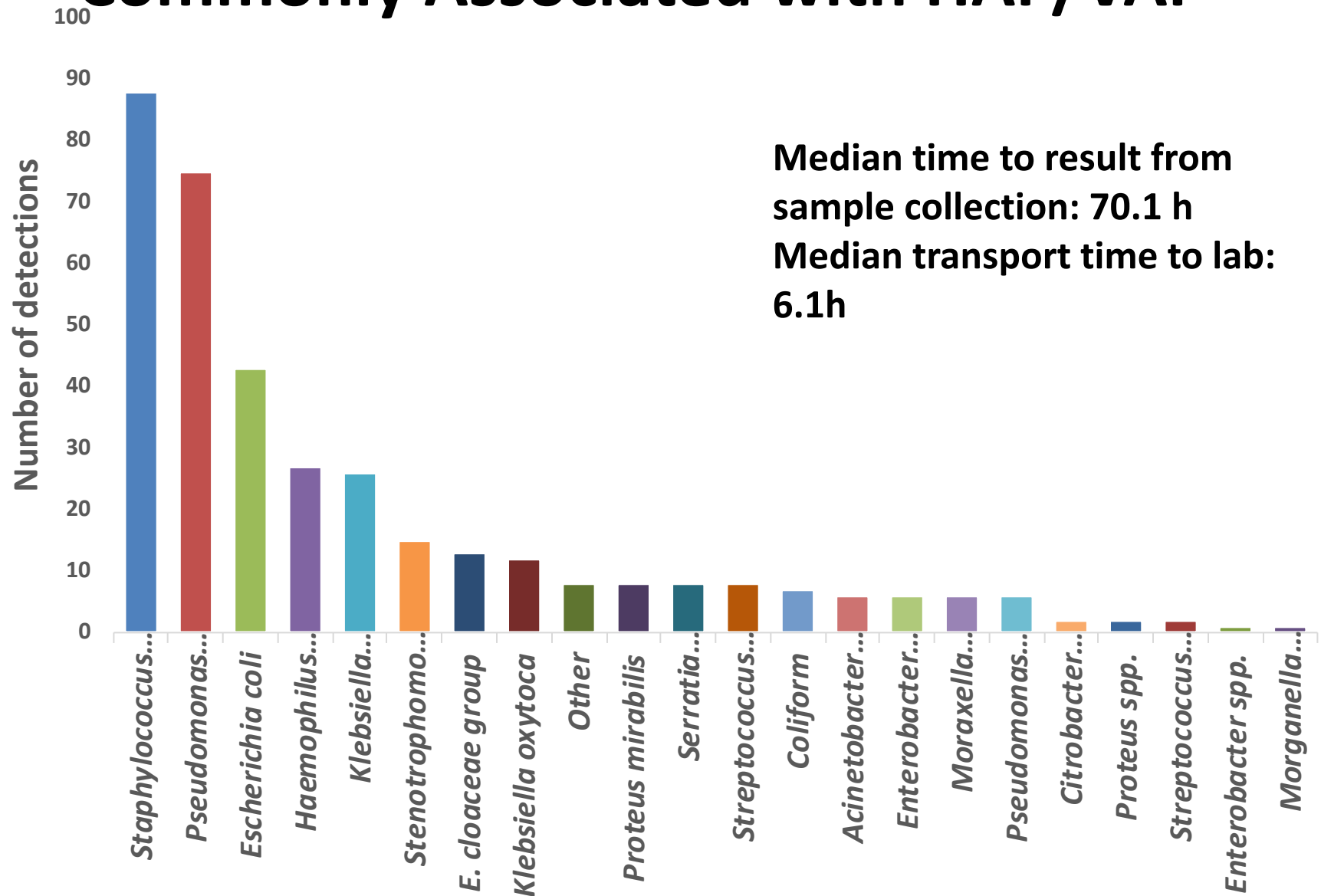
ARTICLES

<https://doi.org/10.1038/s41587-019-0156-5>

Nanopore metagenomics enables rapid clinical diagnosis of bacterial lower respiratory infection

Themoula Charalampous^{1,8}, Gemma L. Kay^{1,2,8}, Hollian Richardson^{1,8}, Alp Aydin², Rossella Baldan^{1,3}, Christopher Jeanes⁴, Duncan Rae⁴, Sara Grundy⁴, Daniel J. Turner⁵, John Wain^{1,2}, Richard M. Leggett⁶, David M. Livermore^{1,7} and Justin O'Grady^{1,2*}

Routine Microbiology finds Organisms Commonly Associated with HAP/VAP



Example Results: UCLH Sample 111 (ETT)

Method	Organism	Resistance Phenotype	Resistance genotype
Routine microbiology	<i>Escherichia coli</i>	Penicillins, amoxy-clav, aztreonam, 2/3/4 gen cephalosporins, co-trimoxazole, trimethoprim	NA
Unyvero	<i>Escherichia coli</i> (+++)	Penicillins, aztreonam, 2/3/4 gen cephalosporins, sulphonamides (predicted)	<i>bla</i> _{TEM-1} <i>sul1</i> <i>bla</i> _{CTX-M} GyrA wt
FilmArray	<i>Escherichia coli</i> (>10 ⁷)	Penicillins, aztreonam, 2/3/4 gen cephalosporins (predicted)	<i>bla</i> _{CTX-M}
MinION	<i>Escherichia coli</i> (90.5% of reads)	Penicillins, aztreonam, 2/3/4 generation cephalosporins, (predicted)	<i>bla</i> _{TEM-1} <i>bla</i> _{CTX-M}

Example Results: RFH Sample 72 (ETT)

Method	Organism	Resistance Phenotype	Resistance genotype
Routine microbiology	Normal Respiratory Flora	NA	NA
Unyvero	<i>E. coli</i> (+++) <i>P. aeruginosa</i> (+++)	Penicillins, fluoroquinolones (predicted)	<i>bla</i> _{TEM-1} GyrA83, GyrA87
FilmArray	<i>E. coli</i> (>10 ⁷) <i>P. aeruginosa</i> (>10 ⁷)	NA	None detected
MinION	<i>E. coli</i> (14.5% reads) <i>P. aeruginosa</i> (43.4% reads)	Penicillins, (predicted)	<i>bla</i> _{TEM-1}

Example Results: CW Sample 5 (ETT)

Method	Organism	Resistance Phenotype	Resistance genotype
Routine microbiology	<i>P. aeruginosa</i> <i>K. pneumoniae</i>	PA: none KP: Gentamicin, trimethoprim	NA
Unyvero	<i>P. aeruginosa</i> (+++) <i>S. maltophila</i> (+)	Penicillins	<i>bla</i> _{SHV}
FilmArray	<i>P. aeruginosa</i> (>10 ⁷) <i>S. agalactiae</i> (10 ⁵)	NA	None detected
MinION	<i>P. aeruginosa</i> (87.8% reads)	NA	None detected

Can MinION Metganomic Sequencing be used to predict AMR?

- Proof of concept study attempting to predict full antimicrobial susceptibility profiles directly from bacteria in INHALE samples
- Compared to full phenotypic susceptibility testing & troubleshooting using PCR, Sanger Sequencing & Illumina Sequencing
- Limited to *E. coli* and *K. pneumoniae* in the first instance

Results: AST phenotype vs MinION prediction

- Successfully identified 7 *E.coli* isolates and 3 *K. pneumoniae* isolates
- AMR Prediction Sensitivity: **71.2%** AMR Prediction Specificity: **98.4%**

Strain	Predicted MinION Phenotype	MinION Genotype	Actual Resistance Phenotype
Sample 1 <i>E. coli</i>	amoxicillin co-trimoxazole gentamicin trimethoprim tobramycin	bla _{TEM-1} aac(3)-II sul1 dfrA12	amoxicillin, co-trimoxazole, gentamicin, piperacillin/tazobactam, co-trimoxazole, tobramycin, trimethoprim, amoxicillin/clavulanate, cefepime
Sample 2 <i>E. coli</i>	amoxicillin streptomycin sulphamethoxazole	bla _{TEM} sul2 strA strB	amoxicillin, streptomycin, sulphamethoxazole, tetracycline
Sample 3 <i>K. Pneumoniae</i>	amoxicillin, amoxicillin/clavulanate, ceftazidime, ciprofloxacin, gentamicin, piperacillin/tazobactam, streptomycin, trimethoprim, suphamethoxazole, co-trimoxazole	bla _{CTX-M} catB3 aac-6'-1b dfrA1 bla _{TEM-1} sul1 straA strB aadA1	amoxicillin, amoxicillin/clavulanate, ceftazidime, ciprofloxacin, gentamicin, piperacillin/tazobactam, streptomycin, trimethoprim, suphamethoxazole, co-trimoxazole, tetracycline

Key:

Agreement

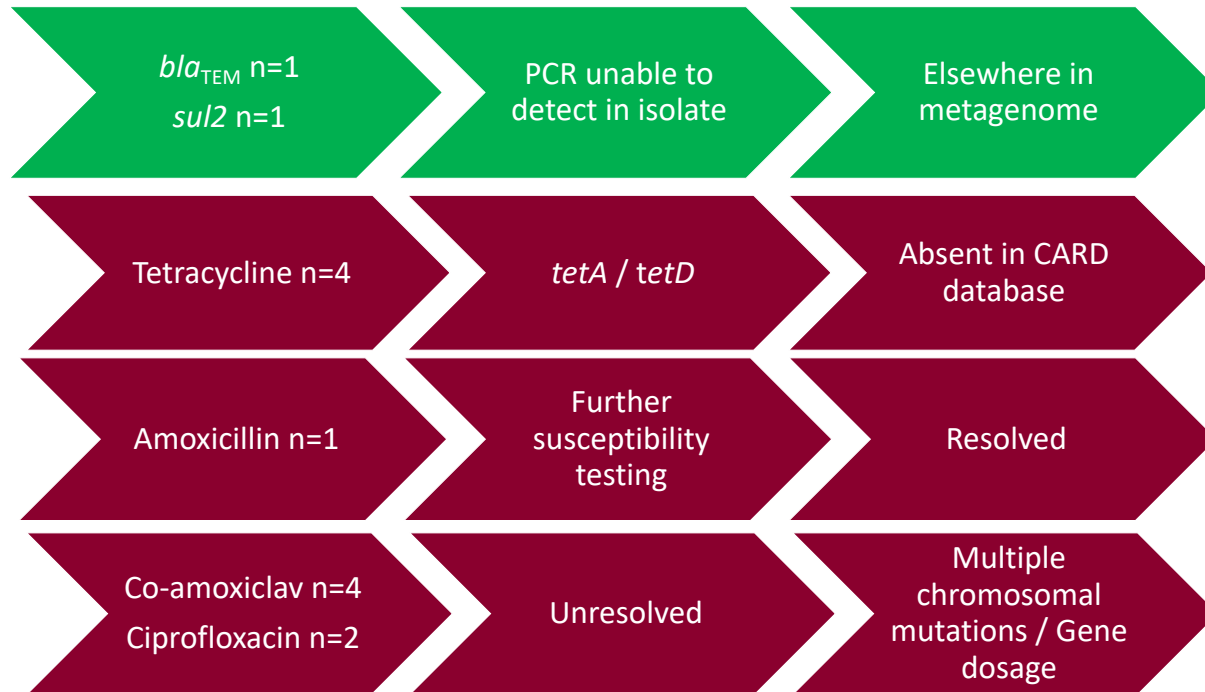
Disagreement

Difficult to Interpret

Discrepancies between culture and MinION

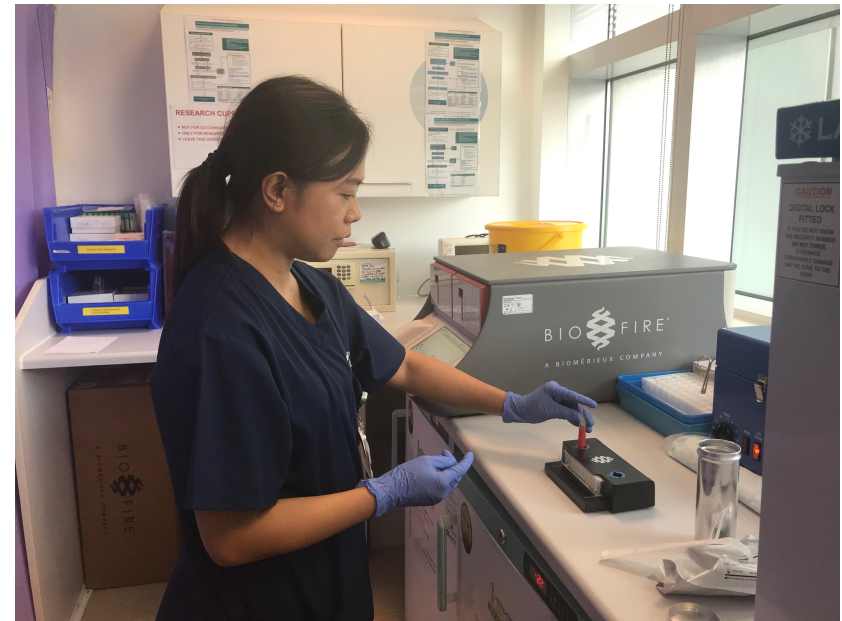
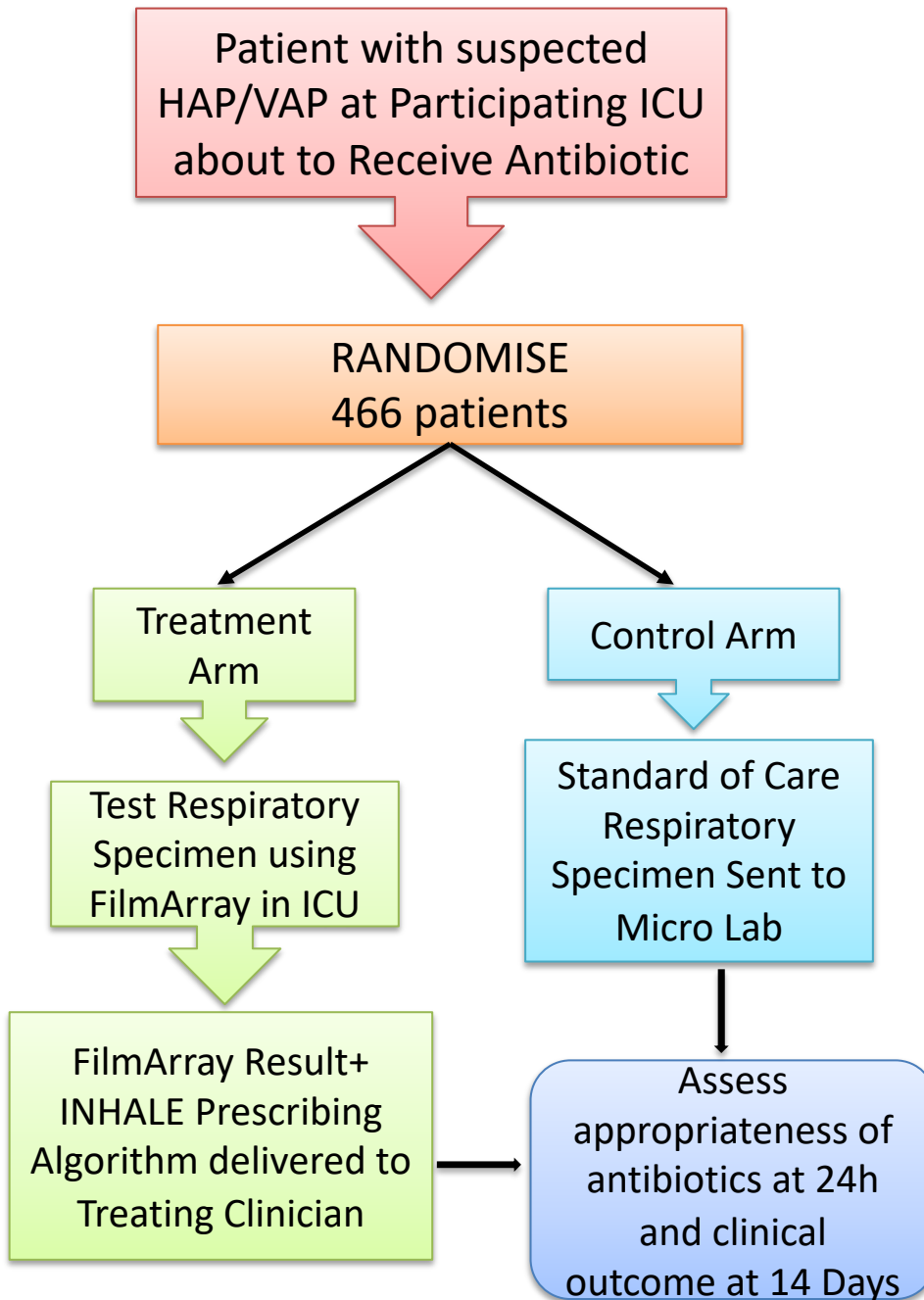
● False Positives

● False Negatives



Demonstrates the **future potential** of MinION sequencing for the **rapid identification** of pathogenic bacteria and predicting resistance phenotypes. Sensitivity should be further improved by **resolving database deficiencies** and using **enhanced bioinformatics**.

WP3 INHALE RCT



FilmArray Torch placed at Point of Care within 12 Critical Care Units

INHALE Prescribing Algorithm

Key

No known allergy to antibiotics

Mild allergy to β -lactams i.e. rash

Severe allergy to β -lactams, i.e. anaphylaxis

Not applicable

Table 2. Recommended treatment for combination of TWO or more organisms are detected by FilmArray

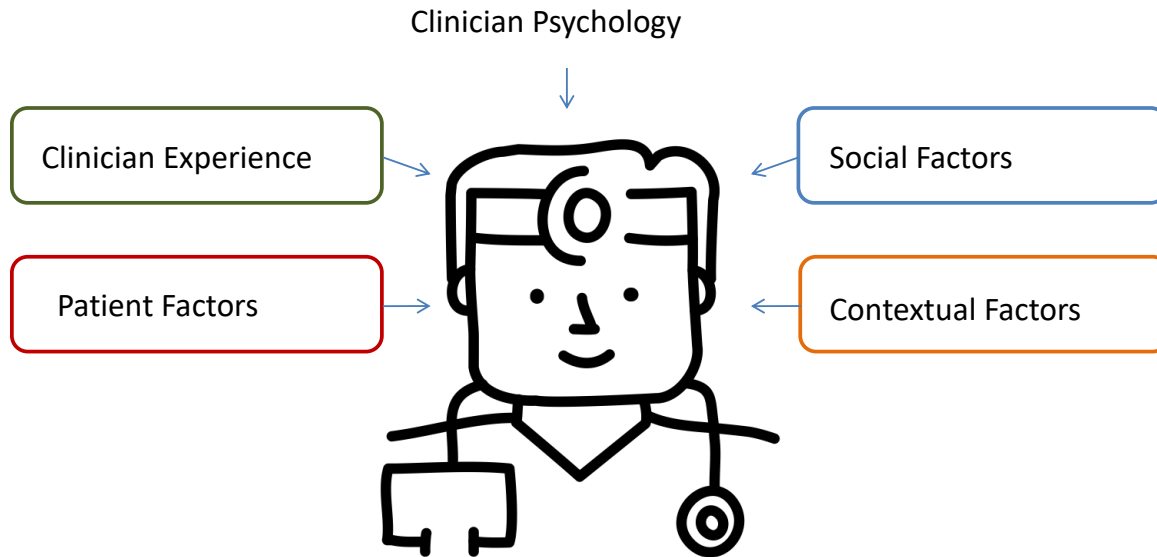
PLEASE READ THIS TABLE FROM LEFT TO RIGHT

Coloured boxes refer to allergy status as in Table 1.

Key: + organism present, - organism absent, \pm either present or absent

First, What combination of bacteria have been found?						Second: Therapy if no resistance genes	Third: if resistance genes found			
<i>A. baumannii</i>	<i>Enterobacteriales:</i> <i>E. aerogenes</i> , <i>E. cloacae</i> , <i>E. coli</i> , <i>K. pneumoniae</i> , <i>K. oxytoca</i> , <i>Proteus</i> sp., <i>S. marcescens</i>	<i>P. aeruginosa</i>	<i>H. influenzae</i> /M. <i>catarrhalis</i>	<i>S. aureus</i>	<i>S. agalactiae</i> , <i>S. pneumoniae</i> or <i>S. pyogenes</i>		<i>mecA</i> /C found	CTX-M found	<i>C. pneumoniae</i> , <i>L. pneumophila</i> OR <i>M. pneumoniae</i>	Carbapen- emase found
Does the mixture include <i>Acinetobacter</i> ? If YES ; stay with this block; if NO, go to next block										
+	Any one or more second organism found					Meropenem ^a	Add Glycopeptide ¹⁰ OR Linezolid	-	Add Macrolide ¹¹ OR Levofloxacin or Ciprofloxacin	Discuss with Micro- biology
+	\pm Any one or more second organism found					Meropenem ^a	Add Glycopeptide ¹⁰ OR Linezolid	-	Add Macrolide ¹¹ OR Levofloxacin or Ciprofloxacin	Discuss with Micro- biology
+	Add Levofloxacin or Ciprofloxacin ⁹	Add Levofloxacin or Ciprofloxacin ⁹	Add Levofloxacin or Ciprofloxacin ⁹	Add Glycopeptide ¹⁰ OR Linezolid	Add Glycopeptide ¹⁰ OR Linezolid	Colistin Combination	Add Glycopeptide ¹⁰ OR Linezolid	Discuss with Micro- biology	Add Macrolide ¹¹ OR Levofloxacin or Ciprofloxacin	Discuss with Micro- biology

WP4: Behavioral Study



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from Noun Project

Consultant: I think time was a big thing because there were fewer people on at night, there is no boss. You're racking around trying to get to see everyone with all the admissions coming in as well. And then you see, oh, he's had a temperature and the nurse is flapping around and keeps phoning me about it, oh, give him some Augmentin and on we go. And perhaps I didn't give it as much thought when I was pressured or rushed.

UCL/UCLH

Dr. Vanya Gant

Mr Alp Aydin

Mr. Dewi Owen

Dr. Zaneeta Dhesi

Dr. Federico Ricciardi

Dr Julie Barber

Dr David Brealey

Ms. Alyssa Pandolfo

Prof. Robert Horne

Industrial Collaborators

bioMerieux Biofire

Oxford Nanopore Technologies

Curetis GmbH

University of East Anglia/Quadram Institute

Prof. David Livermore

Dr. Justin O'Grady

Dr. Rossella Baldan

Dr. Hollian Richardson

Ms. Charlotte Russell

Ms. Juliet High

Mr. Antony Colles

Prof. Ann Marie Swart

The INHALE Study Team & Staff at Participating Hospitals and Laboratories

Funder's Statement

- *This presentation presents independent research funded by the National Institute for Health Research (NIHR) under its Programme Grants for Applied Research Programme (Reference Number RP-PG-0514-20018).*
- *The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health.”*